



## Orthobiologics: Where are we Now?



Steven Sampson<sup>1</sup>, Hunter Vincent<sup>2\*</sup>, Mary A Ambach<sup>3</sup> and Edwin Amirianfar<sup>4</sup>

<sup>1,3</sup>David Geffen School of Medicine at UCLA, The Orthohealing Center, Los Angeles, CA, USA

<sup>2</sup>UC Davis Medical Center, Department of Physical Medicine and Rehabilitation

<sup>4</sup>Western University of Health Sciences, USA

**Submission:** September 09, 2017; **Published:** September 14, 2017

**\*Corresponding author:** Hunter Vincent, UC Davis Medical Center, Department of Physical Medicine and Rehabilitation, Sacramento CA, Dept of PM&R: 4860 Y street, Sacramento, California 95817, USA, Email: [huntervincent35@gmail.com](mailto:huntervincent35@gmail.com)

### Mini Review

Over the last decade, the demand for minimally invasive treatments of musculoskeletal injuries has markedly increased. Recent advances in technology and developments in scientific understanding have created a myriad of new opportunities to meet this demand. One particular area which has experienced a growing volume of research is called orthobiologics. Orthobiologics is defined as any treatment that utilizes the body's native cellular components to promote healing of damaged or diseased tissues [1,2]. Currently, there are 4 generations of orthobiologics including: Hyaluronic acid (HA), platelet-rich plasma (PRP), bone marrow concentrate (BMC), and adipose-derived mesenchymal stem cells (aMSC). Although the current landscape of orthobiologics can be classified by 4 generations, the field as a whole is in the preliminary stages. Continued research and collaboration is needed to expand our understanding of these treatments and shape its future direction.

Hyaluronic acid (HA) is widely considered the first generation of orthobiologics. HA is a naturally occurring protein in humans that has many functions, most importantly acting as an intra articular lubricant, working to reduce friction in synovial joints. Studies have shown overall HA concentration in synovial fluid to decrease with Osteoarthritis[3], with a resultant shift from high molecular weight to low molecular weight variants. In addition, high molecular weight variants have been shown to be superior to low molecular weight variants for chondroprotection in joint osteoarthritis [3]. In theory, the administration of intra articular HA increases the synovial fluid concentration of HA, to possibly prevent symptoms of pain and decrease further joint degeneration [3]. Hyaluronic acid has been shown to bind to cluster of differentiation 44 (CD 44), which inhibits the pro-inflammatory effects of interleukin-1beta, resulting in down regulation of many MMPs associated with cartilage degradation [4,5].

Several clinical trials have shown HA to effectively treat pain associated with OA [6], while also demonstrating a superior

safety profile when compared to continuous NSAID use for pain control in OA [7-9]. In addition, OARSI suggests "good" level of evidence for the treatment of OA with intra articular hyaluronic acid. Although HA has been primarily been used for large joint osteoarthritis, there is a small amount of research investigating its application for spinal pathologies, including facet mediated low back pain, and intradiscal injections of HA-based hydrogels for degenerative disc disease [10,11]. Platelet Rich Plasma (PRP) was first used for open heart surgery in 1987 [12] but has since emerged as the second generation of orthobiologics for musculoskeletal pathology. PRP is extracted from a patient's own blood supply. Venous blood is drawn from the patient and centrifuged in order to separate the blood into multiple layers, including the buffy coat, which contains the largest concentration of platelets. The buffy coat is removed from the processed venous blood and re-injected into various treatment areas [13]. The proposed therapeutic benefit of PRP is based in its ability to stimulate an inflammatory cascade and initiate a healing response through release of growth factors from its alpha granules, including transforming growth factor beta (TGFbeta), vascular endothelial growth factor (VEGF), platelet-derived growth factor (PDGF), and epithelial growth factor (EGF) [13,14]. More recent theories of PRP's mechanism suggest that intra articular application may potentially alter the entire joint environment via the signaling cascade, creating a more advantageous inflammatory environment for healing [15].

Most of the research for PRP consists of heterogenous, small case series, however some larger randomized controlled trials have demonstrated its use in areas such as chronic tendinopathies [16,17] and knee osteoarthritis [18]. In addition, its application for facet mediated low back pain and degenerative disc disease have also started to be researched [19,20]. Some early studies

have also started to examine the combination of PRP with other orthobiologic generations [21-23], as well as various treatment protocol sequences [24], with significant benefits illustrated. However, in all areas of PRP research, uniformity of PRP classification has been lacking, and recent development of PRP classification systems such as the PLRA [25] and MARSPILL [26] have attempted to provide more clarity and standardization across the field, as well as unify research efforts in the future.

The third generation of orthobiologics is bone marrow concentrate (BMC), which consists of a milieu of mesenchymal stem cells (MSCs), hematopoietic cells, platelets, and cytokines noted for possessing anti-inflammatory, immunomodulatory, and chondrogenic properties [27]. While the mechanism is not known, it is thought that BMC either induces differentiation and proliferation of resident stem cells, or possesses innate chondrogenic potential [27]. Procurement of BMC includes aspiration of bone marrow from the patient, usually at the posterior superior iliac crest under fluoroscopic or ultrasound guidance. In a similar manner to PRP, the aspirate is then centrifuged, and specific layers are extracted for injection. Although much of the early research has been mixed, some preliminary studies have demonstrated significant patient safety and efficacy with joint osteoarthritis [28-31]. Research efforts have also been directed towards intradiscal applications in degenerative disc disease, through an ongoing clinical trial called the CASCADE trial, examining the use of BMC with and without HA for discogenic low back pain [32].

The newest and fourth generation of orthobiologics is known as lipoaspirate/adipose derived mesenchymal stem cells (aMSCs). Lipoaspirate is obtained in larger amounts with less invasive techniques via local anesthesia and vacuum-assisted lipectomy to the posterior superior buttock or lateral thigh. In contrast to PRP and BMC, lipoaspirate involves low speed centrifugation or settling of the suctioned adipose tissue for several hours without centrifugation [30]. Similar to BMC, processed lipoaspirate has illustrated chondrogenic, osteogenic, adipogenic, myogenic, and neurogenic differentiation in the presence of certain induction factors [33,34]. Some research has illustrated that aMSCs actually possess larger total numbers of MSCs, however data is mixed as to whether aMSCs have equivalent osteogenic potential as BMC [35,36]. Early research suggests that aMSCs exhibit an anti-inflammatory effect on chondrocytes and synoviocytes in patients with Osteoarthritis [37], however significant research is needed in this generation of orthobiologic.

Orthobiologics is a vastly expanding field within musculoskeletal medicine, currently characterized by 4 generations: hyaluronic acid, platelet rich plasma, bone marrow concentrate, and adipose derived mesenchymal stem cells. Future generations of Orthobiologics are currently being developed, most notably, amniotic tissue as an allogeneic source for mesenchymal stem cells [38,39]. Although applications within

each generation continue to expand, significant research and collaborative efforts are needed to increase our understanding of potential therapeutic benefits and further study the cellular constituents of each orthobiologic.

### References

1. Malanga G, Abdelshahed D, Jayaram P (2016) Orthobiologic Interventions Using Ultrasound Guidance. *Phys Med Rehabil Clin N Am* 27(3): 717-731.
2. Sampson S, Vincent H, Aufiero D (2014) Orthobiologics: A New Generation of Orthopaedics. *Orthopreneur*, p. 1-3.
3. Altman RD, Manjoo A, Fierlinger A, Niazi F, Nicholls M (2015) The mechanism of action for hyaluronic acid treatment in the osteoarthritic knee: a systematic review. *BMC Musculoskelet Disord* 16: 321.
4. Karna E, Milyk W, Surazyński A, Pałka JA (2008) Protective effect of hyaluronic acid on interleukin-1-induced deregulation of  $\beta$ 1-integrin and insulin-like growth factor-I receptor signaling and collagen biosynthesis in cultured human chondrocytes. *Mol Cell Biochem* 308(1-2): 57-64.
5. Waddell DD, Kolomytkin OV, Dunn S, Marino AA (2007) Hyaluronan suppresses IL-1 $\beta$ -induced metalloproteinase activity from synovial tissue. *Clin Orthop Relat Res* 465: 241-248.
6. Moreland LW (2003) Intra-articular hyaluronan (hyaluronic acid) and hylans for the treatment of osteoarthritis: mechanisms of action. *Arthritis Res Ther* 5(2): 54-67.
7. Bannuru RR, Vaysbrot EE, Sullivan MC, McAlindon TE (2014) Relative efficacy of hyaluronic acid in comparison with NSAIDs for knee osteoarthritis: A systematic review and meta-analysis. *Semin Arthritis Rheum* 43(5): 593-599.
8. Bannuru RR, Natov NS, Dasi UR, Schmid CH, McAlindon TE (2011) Therapeutic trajectory following intra-articular hyaluronic acid injection in knee osteoarthritis—meta-analysis. *Osteoarthritis Cartilage* 19(6): 611-619.
9. Colen S, van den Bekerom MP, Mulier M, Haverkamp D (2012) Hyaluronic acid in the treatment of knee osteoarthritis: a systematic review and meta-analysis with emphasis on the efficacy of different products. *BioDrugs* 26(4): 257-268.
10. Cloyd JM, Malhotra NR, Weng L, Chen W, Mauck RL, et al. (2007) Material properties in unconfined compression of human nucleus pulposus, injectable hyaluronic acid-based hydrogels and tissue engineering scaffolds. *Eur Spine J* 16(11): 1892-1898.
11. Tsaryk R, Gloria A, Russo T, Anspach L, De Santis R, et al. (2015) Collagen-low molecular weight hyaluronic acid semi-interpenetrating network loaded with gelatin microspheres for cell and growth factor delivery for nucleus pulposus regeneration. *Acta Biomater* 20: 10-21.
12. Ferrari M, Zia S, Valbonesi M, Henriquet F, Venere G, et al. (1987) A new technique for hemodilution, preparation of autologous platelet-rich plasma and intraoperative blood salvage in cardiac surgery. *Int J Artif Organs* 10(1): 47-50.
13. Sampson S, Gerhardt M, Mandelbaum B (2008) Platelet rich plasma injection grafts for musculoskeletal injuries: a review. *Curr Rev Musculoskelet Med* 1(3-4): 165-174.
14. Alsousou J, Thompson M, Hulley P, Noble A, Willett K (2009) The biology of platelet-rich plasma and its application in trauma and orthopaedic surgery: A review of the literature. *J Bone Joint Surg Br* 91(8): 987-996.
15. Filardo G, Kon E, Roffi A, Di Matteo B, Merli ML (2015) Platelet-rich plasma: why intra-articular? A systematic review of preclinical studies and clinical evidence on PRP for joint degeneration. *Knee Surg Sports Traumatol Arthrosc* 23(9): 2459-2474.

16. Mishra AK, Skrepnik NV, Edwards SG, Jones GL, Sampson S, et al. (2014) Efficacy of platelet-rich plasma for chronic tennis elbow: a double-blind, prospective, multicenter, randomized controlled trial of 230 patients. *Am J Sport Med* 42(2): 463-471.
17. RhaDW, Park GY, Kim YK, Kim MT, Lee SC (2013) Comparison of the therapeutic effects of ultrasound-guided platelet-rich plasma injection and dry needling in rotator cuff disease: a randomized controlled trial. *Clin Rehabil* 27(2): 113-122.
18. Patel S, Dhillon MS, Aggarwal S, Marwaha N, Jain A (2013) Treatment with platelet-rich plasma is more effective than placebo for knee osteoarthritis: a prospective, double-blind, randomized trial. *Am J Sport Med* 41(2): 356-364.
19. Aufiero D, Vincent H, Sampson S, Bodor M (2015) Regenerative Injection Treatment in the Spine: Review and Case Series with Platelet Rich Plasma. *J stem cell Res Rev reports* 1(4).
20. Tolbert G, Roy D, Walker V (2013) Ultrasound Guided Dextrose Prolotherapy and Platelet Rich Plasma Therapy in Chronic Low Back Pain: Three Case Reports. *Int J Phys Med Rehabil* 1: 149.
21. Lana JFSD, Weglein A, Sampson S, Vicente EF, Huber SC, et al. (2016) Randomized controlled trial comparing hyaluronic acid, platelet-rich plasma and the combination of both in the treatment of mild and moderate osteoarthritis of the knee. *J Stem Cells Regen Med* 12(2): 69-78.
22. Koh YG, Kwon OR, Kim YS, Choi YJ (2014) Comparative Outcomes of Open-Wedge High Tibial Osteotomy with Platelet-Rich Plasma Alone or in Combination With Mesenchymal Stem Cell Treatment: A Prospective Study. *Arthroscopy* 30(11): 1453-1460.
23. Pak J, Lee JH, Kartolo WA, Lee SH (2016) Cartilage Regeneration in Human with Adipose Tissue-Derived Stem Cells: Current Status in Clinical Implications. *Biomed Res Int* 2016: 1-12.
24. Sampson S, Smith J, Vincent H, Aufiero D, Zall M, et al. (2016) Intra-articular bone marrow concentrate injection protocol: short-term efficacy in osteoarthritis. *Regen Med* 11(6): 511-520.
25. Mautner K, Malanga GA, Smith J, Shiple B, Ibrahim V, et al. (2015) A Call for a Standard Classification System for Future Biologic Research: The Rationale for New PRP Nomenclature. *PMR* 7(4 Suppl): S53-9.
26. Lana JFSD, Purita J, Paulus C, Huber SC, Rodrigues BL, et al. (2017) Contributions for classification of platelet rich plasma - proposal of a new classification: MARSPILL. *Regen Med* 12(5): 565-574.
27. Sampson S, Botto-van Bemden A, Aufiero D (2013) Autologous bone marrow concentrate: review and application of a novel intra-articular orthobiologic for cartilage disease. *Phys Sport* 41(3): 7-18.
28. Sampson S, Smith J, Vincent H, Aufiero D, Zall M, et al. (2016) Intra-articular bone marrow concentrate injection protocol: short-term efficacy in osteoarthritis. *Regen Med* 11(6): 511-520.
29. Shapiro SA, Kazmerchak SE, Heckman MG, Zubair AC, O'Connor MI (2016) A Prospective, Single-Blind, Placebo-Controlled Trial of Bone Marrow Aspirate Concentrate for Knee Osteoarthritis. *Am J Sports Med* 45(1): 82-90.
30. Centeno C, Pitts J, Al-Sayegh H, Freeman M (2014) Efficacy of autologous bone marrow concentrate for knee osteoarthritis with and without adipose graft. *Biomed Res Int* 2014: 370621.
31. Centeno CJ, Freeman MD (2014) Percutaneous injection of autologous, culture-expanded mesenchymal stem cells into carpometacarpal hand joints: a case series with an untreated comparison group. *Wien Med Wochenschr* 164(5-6): 83-87.
32. (2016) A prospective, Multicenter, Randomized, Double-blind Single, placebo-controlled study to evaluate the efficacy and safety of a single injection of Rexlemestrocel-L alone or combined with hyaluronic acid (HA). Mesoblast Ltd, Australia.
33. Zuk PA, Zhu M, Mizuno H, Huang J, Futrell JW, et al. (2001) Multilineage cells from human adipose tissue: implications for cell-based therapies. *Tissue Eng* 7(2): 211-228.
34. Rodriguez AM, Elabed, C Amri EZ, Ailhaud G, Dani C (2005) The human adipose tissue is a source of multipotent stem cells. *Biochimie* 87(1): 125-128.
35. Park SH, Sim WY, Min BH, Yang SS, Khademhosseini A, et al. (2012) Chip-based comparison of the osteogenesis of human bone marrow- and adipose tissue-derived mesenchymal stem cells under mechanical stimulation. *PLoS One* 7(9): e46689.
36. Hung BP, Hutton DL, Kozielski KL, Bishop CJ, Naved B, et al. (2015) Platelet-Derived Growth Factor BB Enhances Osteogenesis of Adipose-Derived But Not Bone Marrow-Derived Mesenchymal Stromal/Stem Cells. *Stem Cells* 33(9): 2773-2784.
37. Manferdini C, Maumus M, Gabusi E, Piacentini A, Filardo G, et al. (2013) Adipose-derived mesenchymal stem cells exert antiinflammatory effects on chondrocytes and synoviocytes from osteoarthritis patients through prostaglandin E2. *Arthritis Rheum* 65(5): 1271-1281.
38. He Q, Li Q, Chen B, Wang Z (2002) Repair of flexor tendon defects of rabbit with tissue engineering method. *Chinese J Traumatol Zhonghua Chuang shang za zhi* 5(4): 200-208.
39. Willett NJ, Thote T, Lin ASP, Moran S, Raji Y, et al. (2014) Intra-articular injection of micronized dehydrated human amnion/chorion membrane attenuates osteoarthritis development. *Arthritis Res Ther* 16(1): R47.



This work is licensed under Creative Commons Attribution 4.0 License

Your next submission with Juniper Publishers will reach you the below assets

- Quality Editorial service
- Swift Peer Review
- Reprints availability
- E-prints Service
- Manuscript Podcast for convenient understanding
- Global attainment for your research
- Manuscript accessibility in different formats ( Pdf, E-pub, Full Text, Audio)
- Unceasing customer service

Track the below URL for one-step submission

<https://juniperpublishers.com/online-submission.php>